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## White Paper

# Validation of Vena8 Endothelial+™ Biochip

Primary HUVEC - Monocyte (THP-1)  
adhesion assay using Cellix  
VenaFlux™ platform

### Abstract

Culture Primary HUVEC in **Vena8 Endothelial+™** biochips and infuse THP-1 monocytic leukemia cells over primary HUVEC cultured in the channels of five **Vena8 Endothelial+™** biochips at a shear stress of 0.5 dynes/cm<sup>2</sup>.



# Validation of Vena8 Endothelial+™ biochip by Primary HUVEC - Monocyte (THP-1) adhesion assay using Cellix Venaflux™ platform

## INTRODUCTION

Cellix Ltd. has developed a novel Microfluidic Platform consisting of a PC-controlled Nanopump with microfluidic biochips and DucoCell™ analysis software. The Nanopump enables very accurate flow rates to be achieved which are more reproducible and consistent compared to anything currently available. Importantly, flow rates are extremely low ( $5 \text{ pL min}^{-1}$  to  $10 \text{ } \mu\text{L min}^{-1}$ ) and the shear stress levels that the pump can mimic (up to  $450 \text{ dynes cm}^{-2}$ ) are equivalent to those found in blood vessels *in vivo*.

**Vena8 Endothelial+™** Biochip contains 8 parallel enclosed microcapillaries for culturing primary endothelial cells and continuous flow cell based assays. Primary cell monolayer is obtained in less than 3 hrs in the channels which permits the user to perform quick experiments in a single day. Wide range of primary cells can be cultured in **Vena8 Endothelial+™** biochip. For example HUVEC, HAEC, HMVEC etc. Primary endothelial cells are cultured and cell suspensions may then be injected using the **Mirus™ Nanopump** which supports a range of shear stresses / shear flow rates for dynamic flow based assays.

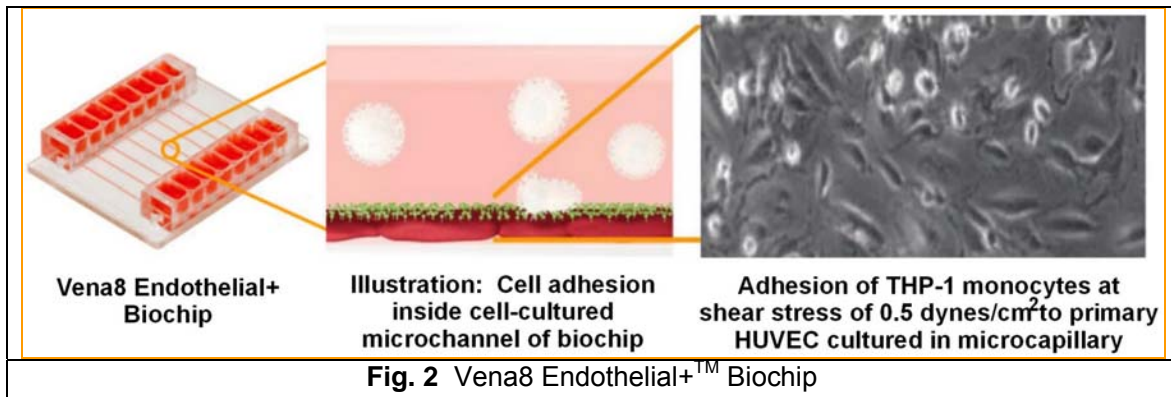
## OBJECTIVE

To culture Primary HUVEC in **Vena8 Endothelial+™** biochips and infuse THP-1 monocytic leukemia cells on primary HUVEC cultured in the channels of five **Vena8 Endothelial+™** biochips at a shear stress of  $0.5 \text{ dynes/cm}^2$ . Specifically, to validate the biochip by estimating channel to channel and biochip to biochip variation by estimating number of THP-1 cells adhered to stimulated Primary HUVEC.



Fig.1 Cellix VenaFlux™ Platform

## Vena8 Endothelial+™ Biochip



Technical Specifications	
Number of channels per biochip	8
Volume of each channel	2.69 $\mu$ L
Depth of channel	120 $\mu$ m / 0.12mm
Width of channel	800 $\mu$ m / 0.8mm
Length of channel	28mm
Dead volume at input port	0.1 $\mu$ L
Vena8 biochip dimensions	800 $\mu$ m (W) x 120 $\mu$ m (D) x 28 mm (L)
Thickness of bottom substrate	0.5mm

Materials	
Adhesion protein	<ul style="list-style-type: none"> <li>Laminin (Sigma L6274)</li> </ul>
Stimulation protein	<ul style="list-style-type: none"> <li>rhTNF-<math>\alpha</math> (Cat.No. 210-TA, R&amp;D Systems)</li> </ul>
<b>Cell line information:</b>	
Primary Cells	<ul style="list-style-type: none"> <li>HUVEC (Human Umbilical Vein Endothelial Cells)</li> </ul>
Growth Properties	<ul style="list-style-type: none"> <li>Adherent</li> </ul>
Organ	<ul style="list-style-type: none"> <li>Human Umbilical Vein</li> </ul>
<b>Cell maintenance</b>	
	<ul style="list-style-type: none"> <li>Endothelial Medium Kit (Promocell C-22110)</li> </ul>
	<ul style="list-style-type: none"> <li>Detach Kit -30 (Promocell C-41200)</li> </ul>
	<ul style="list-style-type: none"> <li>Accutase Solution (Promocell C-41310)</li> </ul>
Cell line	<ul style="list-style-type: none"> <li>THP-1</li> </ul>
Growth Properties	<ul style="list-style-type: none"> <li>Suspension</li> </ul>
Organism	<ul style="list-style-type: none"> <li><i>Homo sapiens</i> (Human)</li> </ul>
Organ	<ul style="list-style-type: none"> <li>Peripheral blood</li> </ul>
Disease	<ul style="list-style-type: none"> <li>Acute monocytic leukemia</li> </ul>
<b>Cell maintenance</b>	
	<ul style="list-style-type: none"> <li>RPMI 1640 (Gibco 31870)</li> </ul>
	<ul style="list-style-type: none"> <li>FBS-10%</li> </ul>
	<ul style="list-style-type: none"> <li>2mM L-glutamine, 100<math>\mu</math>g/ml Penicillin/Streptomycin (Sigma G6784)</li> </ul>

## Method

### i) Stimulation of Primary HUVEC

Primary HUVEC (90% confluent) were stimulated with 10ng/mL rhTNF- $\alpha$  overnight (16-18 hrs) in T75 cm<sup>2</sup> flask.

### ii) Coating Vena8 Endothelial+™ biochips

**Vena8 Endothelial+™** biochip was kept under UV for 20-30mins and all the channels were coated with 12 $\mu$ L of 100 $\mu$ g/mL Laminin (Sigma, Cat.no L-6274). The biochip was then kept in an opened humidified chamber in CO<sub>2</sub> incubator for 1-1.5 hrs.

### iii) Preparation of primary cells

Primary HUVEC were maintained in the recommended growth medium. Cell suspension of  $1.5 \times 10^6$  per 100 $\mu$ L was prepared for seeding in the channels.

### iii) Cell Seeding in Vena8 Endothelial+™ biochip

After the incubation period, 5  $\mu$ L of  $1.5 \times 10^6$  per 100 $\mu$ L of Primary endothelial cells were gently added into each channel of **Vena8 Endothelial+™** biochip. Biochip was kept in the CO<sub>2</sub> incubator for 15 – 20 minutes. The biochip was observed under the microscope and all the reservoirs were topped up with 40 $\mu$ L of media. The biochip was kept for a further 1.5 to 2 hrs in the CO<sub>2</sub> incubator to obtain a Primary cell monolayer inside the channels of the **Vena8 Endothelial+™** biochip.

### iv) Preparation of Cell line

THP1 cells were maintained in the recommended growth media. Cell suspension of  $5 \times 10^6$ /mL cells was prepared for the experiment.

### v) Adhesion Assay

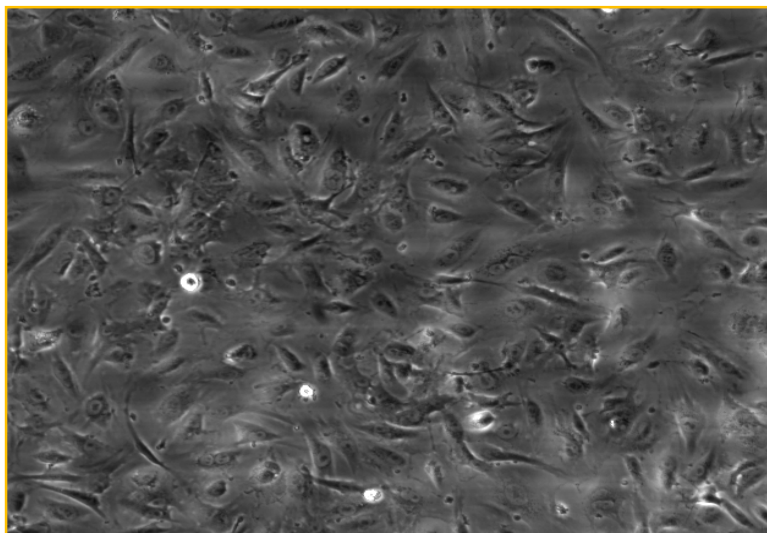
**VenaFlux™ assay software** was used to perform experiments. After the incubation period, THP-1 cells were infused over the HUVEC monolayer inside the channels at a shear stress of 0.5 dynes/cm<sup>2</sup> using the **Mirus Nanopump 2.0**. The THP-1 cells were perfused for 3 mins per channel. Images were acquired at positions 3, 4 and 5 after 2mins 40secs of perfusion in every channel. 10 images per position were captured using a high definition **QImaging** camera with the help of the **Marzhauser IM** series motorized stage. Five **Vena8 Endothelial+™** biochips were used for the assay. All the experiments were performed at 37<sup>o</sup>C using **Oko-lab's Microscope Cage incubator**.

vi) **Image Analysis**

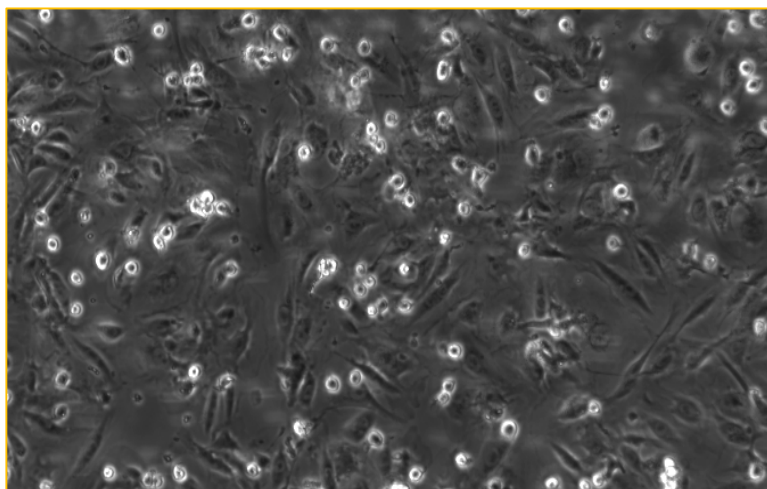
The adhered THP-1 cells on Primary HUVEC were counted using the **Duocell** application software and **Image Pro Analyser 7.0**. The data was exported to Excel for calculations and interpretations

**RESULTS**

An average of **100** cells (THP-1) adhered on Primary HUVEC monolayer per frame. The channel to channel variation was less than **10%** and chip to chip variation was **4%**.



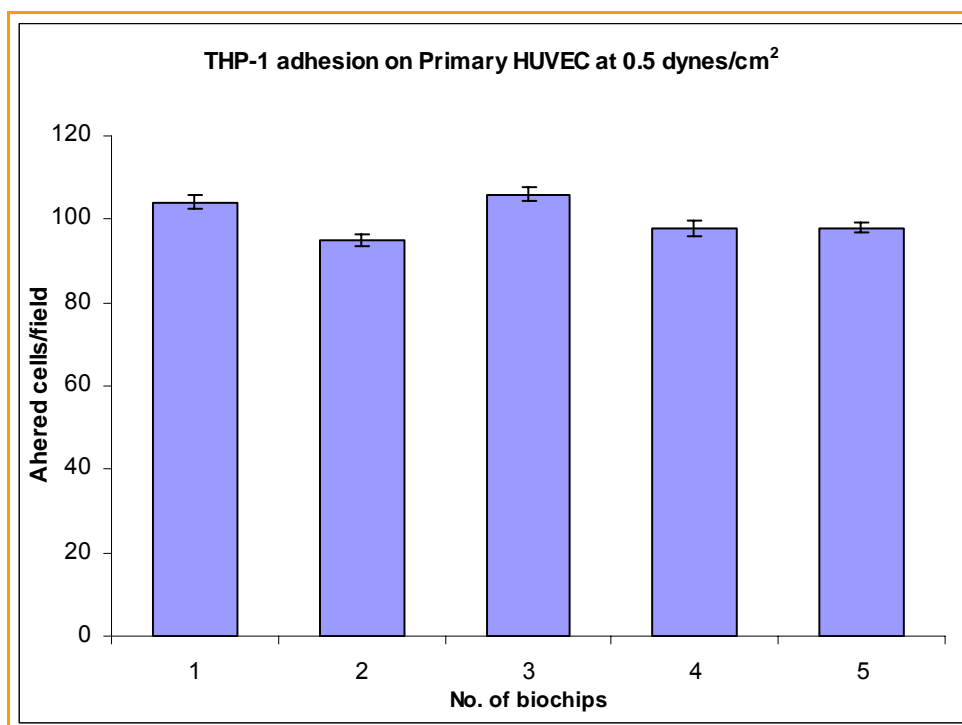
**Fig.3** Primary HUVEC cultured inside Vena8 Endothelial+<sup>1M</sup> biochip channels.



**Fig.4** Adhesion of THP-1 monocytes at a shear stress of 0.5 dyne/cm<sup>2</sup> to 10ng/ml rhTNF- $\alpha$  stimulated Primary HUVEC cultured in channel.

## Results

Channel to Channel analysis					Chip to Chip analysis			
Chip No	Av. No. of Cells (THP1) / 8 channels	STDEV	%CV	SEM	Av. No. of Cells	STDEV	%CV	SEM
1	104	7.76	7.4	1.6	100	4.41	4	0.4
2	95	7.98	8.4	1.6				
3	106	7.59	7.2	1.5				
4	98	8.81	9.0	1.9				
5	98	5.57	5.7	1.2				



**Fig.5** THP-1 cell adhesion to 10ng/mL rhTNF- $\alpha$  stimulated primary HUVEC at a constant shear stress of 0.5 dynes/cm<sup>2</sup> on **Vena8 Endothelial+**<sup>TM</sup> biochip.

### Acknowledgement

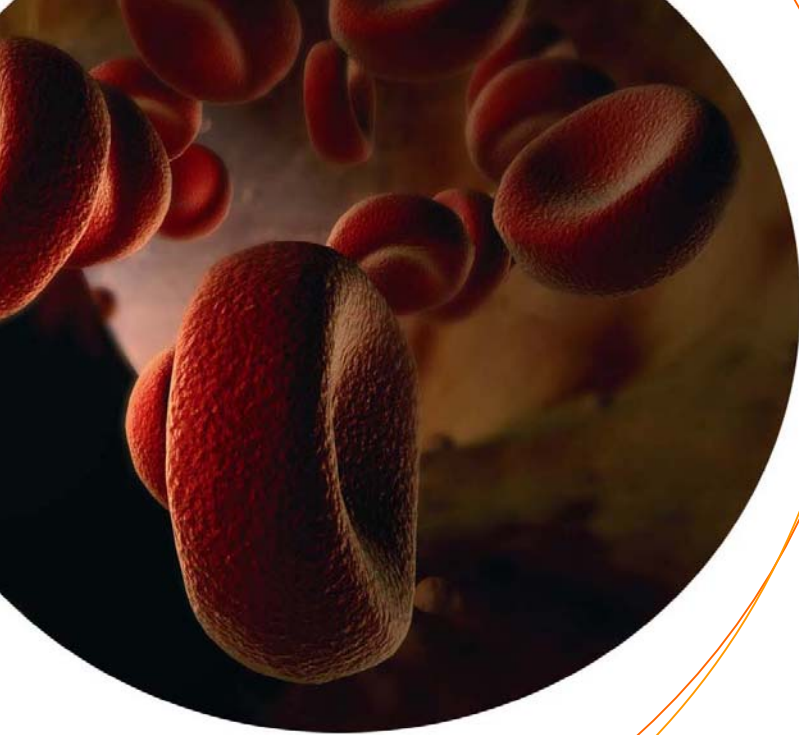
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